

## Renealtins A and B, New Diarylheptanoids with a Tetrahydrofuran Ring from the Seeds of *Renealmia exaltata*

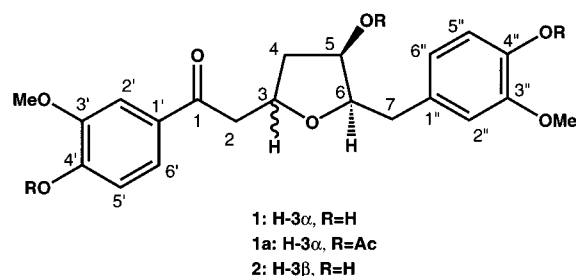
Mitsuhiro Sekiguchi,<sup>†</sup> Hideyuki Shigemori,<sup>†</sup> Ayumi Ohsaki,<sup>\*,‡</sup> and Jun'ichi Kobayashi<sup>\*,†</sup>

Graduate School of Pharmaceutical Sciences, Hokkaido University, Sapporo 060-0812, Japan, and Institute of Biomaterials and Bioengineering, Tokyo Medical and Dental University, Tokyo 101-0062, Japan

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Two new diarylheptanoids, renealtins A (**1**) and B (**2**), have been isolated from the seeds of the Brazilian medicinal plant *Renealmia exaltata* ("Pacová-catinga", Zingiberaceae), and their structures were elucidated by spectroscopic data. Renealtins A (**1**) and B (**2**) are the first example of naturally occurring diarylheptanoids containing a tetrahydrofuran ring.

Brazilian medicinal plants have proven to be a rich source of compounds that might be useful for the development of new pharmaceutical agents.<sup>1</sup> In our search for structurally unique compounds from Brazilian medicinal plants, labdane-derived diterpenoids<sup>2</sup> and nitrogen-containing clerodane diterpenoids<sup>3,4</sup> have been isolated from the leaves of *Echinodorus macrophyllus*. Recent investigations on extracts from the seeds of the Brazilian medicinal plant *Renealmia exaltata* ("Pacová-catinga") led to the isolation of three new labdane diterpenoids, paco-vatinins A–C.<sup>5</sup> Further investigation on extracts from the seeds of *R. exaltata* resulted in the isolation of renealtins A (**1**) and B (**2**), two new diarylheptanoids containing a tetrahydrofuran ring. In this paper we describe the isolation and structure elucidation of **1** and **2**.



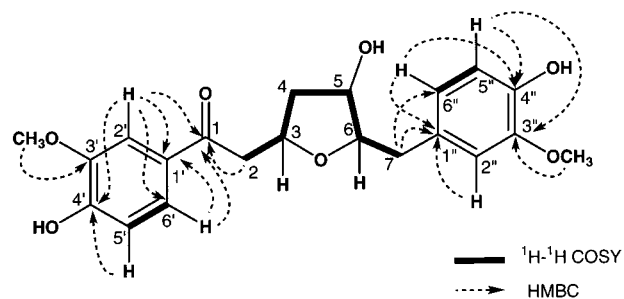
The seeds of *R. exaltata* L.f. (Zingiberaceae) were extracted with MeOH. The MeOH extracts were partitioned between hexane and 90% MeOH, and the MeOH layer was partitioned between EtOAc and H<sub>2</sub>O. The EtOAc-soluble portions were subjected to silica gel column chromatography (CHCl<sub>3</sub>–MeOH, 4:1) followed by reversed-phase C<sub>18</sub> column chromatography (MeOH–H<sub>2</sub>O, 1:1) and then reversed-phase C<sub>18</sub> HPLC (MeOH–H<sub>2</sub>O, 1:1) to afford renealtins A (**1**, 0.000095%) and B (**2**, 0.000095%).

The molecular formula, C<sub>21</sub>H<sub>24</sub>O<sub>7</sub>, of renealtin A (**1**) was established by HRFABMS [*m/z* 411.1420 (M + Na)<sup>+</sup>,  $\Delta$  0.0 mmu]. The IR spectrum suggested the presence of hydroxy (3423 cm<sup>-1</sup>) and unsaturated ketone (1629 cm<sup>-1</sup>) groups, while UV absorptions at 305 and 279 nm implied that **1** possessed a conjugated benzene ring. The gross structure of **1** was deduced from detailed analysis of <sup>1</sup>H and <sup>13</sup>C NMR

**Table 1.** <sup>1</sup>H and <sup>13</sup>C NMR Data of Renealtins A (**1**) and B (**2**) in CD<sub>3</sub>OD

position	<b>1</b>		<b>2</b>	
	<sup>1</sup> H <sup>a</sup>	<sup>13</sup> C <sup>a</sup>	<sup>1</sup> H <sup>a</sup>	<sup>13</sup> C <sup>a</sup>
<b>1</b>		198.36		199.83
2a	3.55 (dd, 6.9,15.6)	48.86	3.24 (m)	45.13
2b	3.10 (dd, 5.8,15.6)		2.94 (dd, 5.8,15.7)	
3	4.33 (m)	75.81	4.73 (m)	75.56
4a	2.53 (m)	41.29	2.18 (m)	38.45
4b	1.78 (m)		1.89 (m)	
5	4.16 (m)	73.10	4.13 (m)	72.86
6a	3.73 (m)	86.00	3.97 (m)	84.83
7a	2.87 (dd, 6.0,13.7)	34.69	2.79 (dd, 6.0,13.7)	35.55
7b	2.81 (dd, 7.2,13.7)		2.70 (dd, 7.2,13.7)	
1'		130.11		130.10
2'	7.58 (d, 1.9)	110.48	7.47 (d, 1.9)	111.62
3'		147.75		148.82
4'		152.56		153.80
5'	6.88 (dd, 8.2)	114.04	6.81 (d, 8.2)	115.33
6'	7.63 (dd, 8.2,1.9)	123.89	7.54 (d, 8.2,1.9)	124.64
1''		131.10		132.66
2''	6.80 (m)	112.24	6.80 (d, 1.9)	113.72
3''		147.55		148.46
4''		144.88		145.92
5''	6.66 (brd)	114.40	6.66 (brd)	115.50
6''	6.66 (brd)	121.23	6.66 (brd)	122.06
3'-OMe	3.90 (s)	55.44	3.80 (s)	56.10
3''-OMe	3.72 (s)	55.55	3.72 (s)	56.10

<sup>a</sup>  $\delta$  in ppm.



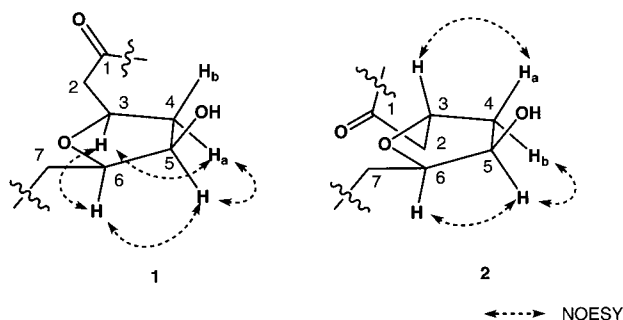
**Figure 1.** Selected 2D NMR data of renealtin A (**1**).

data (Table 1) aided with 2D NMR experiments (<sup>1</sup>H–<sup>1</sup>H COSY, HMQC, and HMBC) (Figure 1). The <sup>13</sup>C NMR data indicated that the molecule possessed one ketone, two benzene rings, three oxymethines, two methoxy groups, and three methylenes. Since nine of 10 unsaturations were accounted for, **1** was inferred to contain one more ring. The <sup>1</sup>H–<sup>1</sup>H COSY (Figure 1) spectrum indicated connectivities of C-2 to C-7, C-5' to C-6', and C-5'' to C-6''. The coupling

\* To whom correspondence should be addressed. Tel: +81-11-706-4985. Fax: +81-11-706-4989. E-mail: jkobay@pharm.hokudai.ac.jp. (J.K.). Tel: +81-3-5280-8153. Fax: +81-3-5280-8005. E-mail: a-ohsaki.fm@tmd.ac.jp. (A.O.).

<sup>†</sup> Hokkaido University.

<sup>‡</sup> Tokyo Medical and Dental University.



**Figure 2.** Selected NOESY correlations and relative stereochemistry of renealtins A (**1**) and B (**2**).

constants and chemical shifts (Table 1) of H-2', H-5', H-6', H-2'', H-5'', and H-6'' suggested the presence of a 3'-methoxy-4'-hydroxybenzene ring and a 3''-methoxy-4''-hydroxybenzene ring. HMBC correlations of H<sub>2</sub>-2 and H-2' to C-1 ( $\delta_C$  198.36) and H<sub>2</sub>-7 to C-1' ( $\delta_C$  131.11) revealed connectivities between C-1' and C-2 through a ketone carbonyl (C-1) and between C-1'' and C-7, respectively. Treatment of **1** with acetic anhydride in pyridine afforded the triacetate (**1a**), judging from its <sup>1</sup>H NMR and MS spectra. The proton signals at C-3 and C-6 in **1a** were not shifted as compared with those of **1**, indicating that **1a** possessed a tetrahydrofuran ring (C-3–C-6 and O-3) with a hydroxy group at C-5. Thus, the gross structure of renealtin A was elucidated to be **1**. NOESY correlations (Figure 2) of H-3/H-4a, H-3/H-6, H-4a/H-5, and H-5/H-6 indicated *syn* relationships both between H-3 and H-6 and between H-5 and H-6. Therefore, the relative stereochemistry of renealtin A (**1**) was assigned as shown in Figure 2.

Renealtin B (**2**) showed the molecular ion peak at *m/z* 411 (M + Na)<sup>+</sup> in the FABMS. HRFABMS analysis revealed the molecular formula to be C<sub>21</sub>H<sub>24</sub>O<sub>7</sub> [*m/z* 411.1408 (M + Na)<sup>+</sup>,  $\Delta$  -1.2 mmu], which was the same as that of renealtin A (**1**). <sup>1</sup>H and <sup>13</sup>C NMR data (Table 1) of **2** were quite similar to those of **1**. Detailed analysis of 2D NMR data (<sup>1</sup>H–<sup>1</sup>H COSY, HMQC, and HMBC) suggested that the gross structure of **2** was the same as that of **1**. NOESY correlations (Figure 2) of H-3/H-4a, H-4b/H-5, and H-5/H-6 indicated *anti* and *syn* relationships between H-3 and H-6 and between H-5 and H-6, respectively. Thus, renealtin B (**2**) was assigned to be the epimer at C-3 of renealtin A (**1**).

Renealtins A (**1**) and B (**2**) are the first example of diarylheptanoids containing a tetrahydrofuran ring, although some diarylheptanoids possessing a tetrahydropyran ring have been reported from *Alpinia blepharocalyx*<sup>6</sup> and *Zingiber officinale*.<sup>7</sup>

## Experimental Section

**General Experimental Procedures.** Optical rotations were determined on a JASCO P-1030 polarimeter. UV and IR spectra were obtained on JASCO Ubest-35 and JASCO FT/IR-230 spectrometers, respectively. <sup>1</sup>H and <sup>13</sup>C NMR spectra were recorded on a Bruker ARX-500 spectrometer. The 3.35 and 49.8 ppm resonances of residual CD<sub>3</sub>OD were used as internal references for <sup>1</sup>H and <sup>13</sup>C NMR spectra, respectively. FAB mass spectra were measured on a JEOL HX-110 spectrometer using a glycerol matrix.

**Plant Material.** The seeds of *Renealmia exaltata* ("Pacovátinga", Zingiberaceae) were purchased in São Paulo, Brazil,

in March 2000. The plant was identified by Dr. G. Hashimoto (Centro de Pesquisas de História Natural, São Paulo, Brazil), and a voucher specimen has been deposited at the Institute of Biomaterials and Bioengineering, Tokyo Medical and Dental University.

**Extraction and Separation.** The seeds (420 g) were extracted with MeOH (500 mL × 3), and the extracts were partitioned between hexane (500 mL × 3) and 90% MeOH (500 mL). The MeOH layer was partitioned with EtOAc (500 mL × 3) and H<sub>2</sub>O (50 mL). The EtOAc-soluble portions (4.2 g) were subjected to a silica gel column (CHCl<sub>3</sub>–MeOH, 4:1) followed by a C<sub>18</sub> column (Cosmosil ODS, MeOH–H<sub>2</sub>O, 1:1) to give a fraction (135 mg). The fraction was subjected to a silica gel column (hexane–acetone, 1:1) followed by reversed-phase HPLC [Develosil ODS HG-5, Nomura Chemical, 1 × 25 cm; eluent, MeOH–H<sub>2</sub>O (1:1); flow rate 2.5 mL/min] to give renealtins A (**1**, *t<sub>R</sub>* 12.8 min, 0.4 mg) and B (**2**, *t<sub>R</sub>* 13.6 min, 0.4 mg).

**Renealtin A (1):** yellow amorphous solid; [ $\alpha$ ]<sub>D</sub><sup>25</sup> +22.4° (c 0.40, MeOH); UV (MeOH)  $\lambda_{max}$  (log  $\epsilon$ ) 305 (3.82), 279 (3.99), and 228 (4.25) nm; IR (KBr)  $\nu_{max}$  3423, 2925, and 1629 cm<sup>-1</sup>; <sup>1</sup>H and <sup>13</sup>C NMR (Table 1); HMBC (H/C) 2/1 2/3 7/6 7/1'' 7/2'' 7/6'' 2/1' 2/1'' 2/3' 2/4' 5/4' 5/1' 6/1' 6/1'' 6/2' 6/4' 3'-OMe/3' 2''/1'' 5''/3'' 5''/4'' 6''/1'' 6''/4'' 3''-OMe/3''; NOESY (H/H) 2a/3 2a/2' 2a/6' 2b/3 2b/2' 2b/6' 3/4a 3/6 4a/5 5/6 7/2'' 7/6'' 5/6' 2/1' 3'-OMe 2''/3''-OMe; FABMS *m/z* 411 (M + Na)<sup>+</sup>; HRFABMS *m/z* 411.1420 (M + Na)<sup>+</sup> (calcd for C<sub>21</sub>H<sub>24</sub>O<sub>7</sub>Na, 411.1420).

**Renealtin B (2):** yellow amorphous solid; [ $\alpha$ ]<sub>D</sub><sup>25</sup> +73.5° (c 0.15, MeOH); UV (MeOH)  $\lambda_{max}$  (log  $\epsilon$ ) 304 (3.88), 279 (4.04), and 228 (4.29) nm; IR (KBr)  $\nu_{max}$  3430, 2925, and 1629 cm<sup>-1</sup>; <sup>1</sup>H and <sup>13</sup>C NMR (Table 1); HMBC (H/C) 2/1 2/3 4/3 4/5 7/5 7/6 7/1'' 7/2'' 7/6'' 2/1' 2/3' 2/4' 2/6' 5/1' 5/3' 6/1' 6/2' 6/4' 2''/1'' 5''/6'' 6''/7 6''/1'' 6''/4''; NOESY (H/H) 2a/6' 2b/6' 3/4a 4b/5 4b/6 5/6 6/7a 6/7b 7a/2'' 7a/6'' 7b/2'' 7b/6'' 5/6' 2/1' 3'-OMe 2''/3''-OMe; FABMS *m/z* 411 (M + Na)<sup>+</sup>; HRFABMS *m/z* 411.1408 (M + Na)<sup>+</sup> (calcd for C<sub>21</sub>H<sub>24</sub>O<sub>7</sub>Na, 411.1420).

**Acetylation of 1.** To a solution of renealtin A (**1**) (0.05 mg) in pyridine (30  $\mu$ L) at room temperature was added Ac<sub>2</sub>O (30  $\mu$ L). The mixture was stirred for 3 h, and the solvent was removed in vacuo to yield **1a** (0.06 mg): <sup>1</sup>H NMR (CD<sub>3</sub>OD)  $\delta_H$  3.49 (1H, m, H-2a), 3.17 (1H, m, H-2b), 4.72 (1H, m, H-3), 2.64 (1H, m, H-4a), 1.78 (1H, m, H-4b), 5.23 (1H, m, H-5), 4.01 (1H, m, H-6), 2.87 (2H, m, H-7), 7.68 (1H, m, H-2'), 7.18 (1H, d, 8.7, H-5'), 7.66 (1H, m, H-6'), 6.85 (1H, m, H-2''), 6.86 (1H, m, H-5''), 6.74 (1H, m, H-6'') 3.88 (3H, s, 3'-OMe), and 3.70 (3H, s, 3''-OMe); FABMS *m/z* 537 (M + Na)<sup>+</sup>.

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